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## ***Data in Brief: Data article***

### ***Data characterizing the chloroplast genomes of extinct and endangered Hawaiian endemic mints (Lamiaceae) and their close relatives***

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#### **Abstract**

These data are presented in support of a plastid phylogenomic analysis of the recent radiation of the Hawaiian endemic mints (Lamiaceae), and their close relatives in the genus *Stachys*, “The quest to resolve recent radiations: Plastid phylogenomics of extinct and endangered Hawaiian endemic mints (Lamiaceae)” [1]. Here we describe the chloroplast genome sequences for 12 mint taxa. Data presented include summaries of gene content and length for these taxa, structural comparison of the mint chloroplast genomes with published sequences from other species in the order Lamiales, and comparisons of variability among three Hawaiian taxa vs. three outgroup taxa. Finally, we provide a list of 108 primer pairs targeting the most variable regions within this group and designed specifically for amplification of DNA extracted from degraded herbarium material.

### Specifications Table

Subject area	<i>Biology, genetics, genomics</i>
More specific subject area	<i>Molecular phylogenetics and evolution</i>
Type of data	<i>Tables and figures</i>
How data was acquired	<i>High-throughput sequencing of contemporary and herbarium samples was conducted on the Illumina HiSeq 2500 and MiSeq platforms, followed by both de novo and reference-guided assemblies, mapping, and functional annotation</i>
Data format	<i>Raw, and analyzed</i>
Experimental factors	<i>De novo assemblies were created using SOAPdenovo and reference-guided assemblies were created using YASRA. Sequences were functionally annotated using DOGMA. SNPs were called and filtered using SAMtools and BCFtools</i>
Experimental features	<i>Data include chloroplast genome gene content, structure, and comparisons of variable loci in a suite of recently diverged species and outgroups</i>
Data source location	<i>Hawaii, North America, South America, Europe, Africa, and Asia</i>
Data accessibility	<i>Data are published with this article</i>

### Value of the data

- These data provide a summary of the characteristics and structure of the chloroplast genomes of several taxa within Lamiaceae, which can be used to increase our understanding of molecular evolution of the chloroplast genome, as well as the evolution of its structure and function.
- A comparison of variable regions in mints can be used to identify rapidly evolving regions in other taxa
- Primer sequences described here can be used to target highly variable regions in closely related taxa

### Data

Raw, demultiplexed sequence reads have been deposited in the NCBI sequence read archive (SRP070171) and full chloroplast genomes for 12 mint taxa have been deposited in GenBank (KU724131-KU724141). Data presented in the text include tables and figures giving information on gene content and variability in these 12 species, as well as comparison of genome structure with other members of Lamiaceae.

### Experimental design, materials and methods

#### *1.1 Samples, library construction, and shotgun sequencing*

We selected 12 Hawaiian mint taxa for shotgun sequencing (five contemporary and seven from herbarium collections ranging up to ~100 years old), of which two extinct species were

represented by two accessions each (see Tables 1 and 2 in [1]). We also sequenced four *Stachys* species, representing both close and more distantly related relatives.

DNA extraction, library construction and shotgun sequencing followed the methods described in [1]. Briefly, approximately 100 mg dried leaf tissue was homogenized using the TissueLyser system (Qiagen), and DNA was extracted using the DNeasy plant mini kit (Qiagen). DNA isolated from herbarium samples was processed separately from contemporary samples using stringent protocols and controls to prevent and detect any potential contamination. For contemporary samples, DNA extracts were sheared to 200 – 600 bp via sonication in a Covaris S220; DNA from herbarium samples is naturally degraded and therefore was not sheared further. Genomic shotgun sequencing libraries were constructed following the standard Illumina Tru-seq protocol for contemporary samples, or the NEBNext Library Prep Mastermix kit (New England Biolabs) for herbarium samples. Libraries were quantified using the PicoGreen High Sensitivity assay and then pooled and sequenced on the Illumina HiSeq and MiSeq platforms. Adapter sequences were trimmed from the reads using the AdapterRemoval software [2]. Assessment of DNA damage in old herbarium specimens was conducted using mapDamage 2.0 [3]. The presence of misincorporations characteristic of damaged DNA molecules typically found in old and degraded samples suggests that the data from herbarium samples are authentic, however, the overall levels of damage were low and within the range expected based on the age of the specimens (see Supplementary Figures 2 and 3 in [1]).

### 1.2 Assembly of the Hawaiian mint reference chloroplast genome

Because no chloroplast genome sequence from a closely related taxon was available at the time this study was conducted, we implemented a combined reference-guided and *de novo* assembly approach [4] to determine the first complete chloroplast genome sequence for a Hawaiian mint. We assembled the sequence for *Stenogyne haliakalae*, an extinct species, as it had the largest number of reads. Briefly, the approach involved conducting both reference-guided assembly in YASRA 2.32 [5] with olive (*Olea europaea*, NC\_013707; [6]) as the reference, as well as *de novo* assembly in SOAPdenovo v1.05 [7]. Assembly methods are described in more detail in [1]. The resulting contigs from both approaches were split into overlapping sequences, and then used as input for a further reference guided-assembly step in YASRA. Gaps between the final contigs were closed using PCR (see [1] for PCR reaction conditions and Supplementary Table 1 of this paper for primer information) and Sanger sequencing in both directions from high-quality DNA extracted from a contemporary sample of *Stenogyne bifida*. This ensured that amplification could be carried out over potentially large gaps, which would not be possible with degraded DNA from the extinct *Stenogyne haliakalae*. Contigs and Sanger sequences were aligned in Sequencher 4.7 (Gene Codes) to create a pseudo-reference sequence [4]. Reads from *Stenogyne haliakalae* were then mapped to the pseudo-reference using BWA v. 0.6.2 [8]. The reference sequence was further refined through Sanger sequencing of areas with low coverage or poor mapping quality (e.g., the border between the inverted repeat and single copy region). Reads were mapped to the final sequence, PCR duplicates were flagged and removed with the MarkDuplicates tool of the Picard command line toolset (<http://picard.sourceforge.net/index.shtml>), and a consensus sequence was called using SAMtools [9].

### 1.3 Assembly of additional mint chloroplast genomes

Complete or nearly complete chloroplast genomes were assembled using similar methods for 11 additional taxa: seven from the endemic Hawaiian mints (two of which were from herbarium samples) and four *Stachys* outgroups (see Tables 1 and 2 in [1]). Since the Hawaiian mints have diverged recently, we used the new chloroplast genome sequence from *Stenogyne haliakalae* as the reference during reference-guided assembly for the remaining Hawaiian taxa. The resulting contigs were aligned to create an interim sequence, and then the reads were mapped to this using BWA and a final consensus sequence called using SAMtools.

Chloroplast genome sequences for the *Stachys* outgroup taxa were assembled in a similar manner. We first assembled the chloroplast genome sequence for *Stachys chamissonis*, as this species is most closely related to the Hawaiian lineage. We conducted independent YASRA runs using *Olea europaea* as the reference, in addition to newly available sequences from *Stenogyne haliakalae*, *Sesamum indicum* (NC\_016433) [10], *Origanum vulgare* (JX880022) [11], and *Salvia miltiorrhiza* (NC\_020431) [12]. The contigs from all five runs were aligned to create an interim sequence. The reads were then mapped to the interim sequence using BWA and a consensus called using SAMtools. Once the *Stachys chamissonis* sequence was assembled we used this as the reference in YASRA for reference guided assembly of both *Stachys coccinea* and *Stachys sylvatica*. For *Stachys byzantina*, the most distantly related outgroup, we performed the initial reference-guided assembly using the sequence from *Stachys chamissonis*, as well as *Olea europaea* and *Sesamum indicum*. The rest of the assembly proceeded as described for the other *Stachys* species.

#### 1.4 Gene content and structure of mint chloroplast genomes

The *Stenogyne haliakalae* reference sequence and sequences from additional species were annotated using a combination of DOGMA [13], tRNAscan-SE [14], and additional manual BLAST searches. The borders of the inverted repeats were identified with the program Inverted Repeats Finder [15].

Overall the chloroplast genome sequences assembled here are very similar to other Lamiales. Table 1 shows the gene content of the *Stenogyne haliakalae* chloroplast genome, which mirrors the gene content of the chloroplast genomes for the 11 additional mint taxa, including the *Stachys* outgroups. Table 2 lists the genes that contain introns (and the number of introns present) in the mint taxa investigated here. Table 3 gives the lengths of the full chloroplast genome for each sequence assembled, as well as the lengths of the inverted repeats and single copy regions.

To compare the genome structure of the Hawaiian and *Stachys* taxa to other taxa in the order Lamiales, we conducted analyses in Mauve 2.3.1 [16] (Figure 1). In the analysis we included *Stenogyne haliakalae*, *Stachys byzantina*, *Ajuga reptans* (NC\_023102), *Andrographis paniculata* (NC\_022451), *Boea hygrometrica* (NC\_016468), *Jasminum nudiflorum* (NC\_008407), *Lindenbergia philippensis* (NC\_022859), *Olea europaea* (NC\_013707), *Origanum vulgare* (JX880022), *Pinguicula ehlersiae* (NC\_023463), *Salvia miltiorrhiza* (NC\_020431), *Schwalbea americana* (NC\_023115), *Sesamum indicum* (NC\_016433), *Tectona grandis* (NC\_020098), and *Utricularia gibba* (NC\_021449). The chloroplast genome sequences of some taxa in this analysis

demonstrated rearrangements and inversions. Therefore, one of the inverted repeats was trimmed off at the coordinates suggested by Inverted Repeats Finder so that homology could be determined with the remaining region. Seed weight was set to 19, the gap opening penalty was set to -200, and the gap extension penalty to -30.

To compare the genome structure of all of the Hawaiian and *Stachys* mints to each other, we conducted a separate analysis in Mauve (Figure 2). The sequences were assumed to be collinear. The seed weight was set to 7, the gap opening penalty was set to -200, and the gap extension penalty to -30.

### 1.5 Variability in the chloroplast genome sequences of Hawaiian mints and *Stachys* outgroups

We investigated variability among the three highest quality chloroplast genome sequences of the Hawaiian mints (*Stenogyne haliakalae*, *Stenogyne bifida*, *Haplostachys haplostachya*), and the three highest quality sequences from the *Stachys* outgroups (*Stachys chamissonis*, *Stachys coccinea*, and *Stachys sylvatica*). These species represent all of the main lineages within our samples, and were sequenced at > 10X depth. We used BWA to map the reads for each of these species onto the *Stenogyne haliakalae* reference genome and used SAMtools and BCFtools to call SNPs with a SNP quality score > 30. Annotations of the *S. haliakalae* reference genome were transferred to the locations of the SNPs. We compared the levels of chloroplast genome diversity among the six genomes by identifying unique, variable positions, which we refer to as potentially informative characters (PICs) [17, 18]. We did not include indels and inversions in this definition, which have been included in other analyses of chloroplast genome variability.

To analyze diversity among the chloroplast genomes, we compared the number of PICs present in 1,000 bp non-overlapping sliding windows across the entire chloroplast genome sequences (Table 4, see also Figure 5 in [1]). We also compared the number of PICs per locus for coding (Table 5, Figure 3a), intron (Table 6, Figure 3b), and intergenic spacer and pseudogene regions (Table 7, Figure 3c). Because this approach does not take into account the length of the locus, very long loci appear to have more PICs than shorter loci. Therefore, we also divided the number of PICs by the total length of the region to give the percent PICs per locus (Tables 5 – 7, Figure 4). However, very short regions may still appear to have a high percentage of variable sites, when in fact only a small number of the sites were variable. To minimize this, we have excluded regions less than 100 bp in length, and for clarity we have also excluded regions that were conserved among all six taxa.

To identify the most variable regions of the mint chloroplast genome for targeted re-sequencing and high resolution phylogenetic analyses, reads from all 15 taxa subjected to shotgun sequencing (including the partial genomes from all of the historical samples, except *Phyllostegia variabilis*) were mapped to the *Stenogyne haliakalae* reference sequence using BWA. SNPs were called with SAMtools and BCFtools, filtering out those with SNP quality < 30. We selected a total of 108 variable loci (see Figure 2 in [1] for diagram of locations) identified from single copy regions, including (1) all the regions that had a variant position among the Hawaiian mints (except where every individual had an alternative allele as compared to the reference sequence) and (2) additional regions that had variant positions among at least two of the *Stachys* species. 100 bp of flanking sequence on either side of the SNPs was retrieved from the reference genome,



and PCR primers were designed using BatchPrimer3 [19], with further manual examination for quality control (e.g. to ensure that primer sequences did not fall into a gap for one of the other taxa). Sequences complementary to the Illumina sequencing adapters were appended to the end of each primer (Table 8) so that sequencing libraries could be prepared directly from the cleaned multiplex PCR products. Overall, these regions represent roughly 20,000 bp of sequence from the chloroplast genome and contain additional variable sites beyond the initial targeted SNP.

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**Table 1. Gene content of the chloroplast genome of *Stenogyne haliakalae* and 11 additional mint species.**

Gene Products	Genes
Photosystem I	psaA, psaB, psaC, psaI, psaJ, ycf3 <sup>[20]</sup> , ycf4 <sup>[20]</sup>
Photosystem II	psbA, psbB, psbC, psbD, psbE, psbF, psbH, psbI, psbJ, psbK, psbL, psbM, psbN, psbT, psbZ/lhbA
Cytochrome b6/f	petA, petB, petD, petG, petL, petN
ATP synthase	atpA, atpB, atpE, atpF, atpH, atpI
Rubisco	rbcL
NADH oxidoreductase	ndhA, ndhB*, ndhC, ndhD, ndhE, ndhF, ndhG, ndhH, ndhI, ndhJ, ndhK
RNA polymerase	rpoA, rpoB, rpoC1, rpoC2
Large subunit ribosomal proteins	rpl2*, rpl14, rpl16, rpl20, rpl22, rpl23*, rpl32, rpl33, rpl36
Small subunit ribosomal proteins	rps2, rps3, rps4, rps7*, rps8, rps11, rps12**, rps14, rps15, rps16, rps18, rps19
Other functions	accD, ccsA, cemA, clpP, matK, infA
Unknown functions	ycf1**, ycf2*, ycf15*
Ribosomal RNAs	rrn23*, rrn16*, rrn5*, rrn4.5*
Transfer RNAs	trnA(UGC)*, trnC(GCA), trnD(GUC), trnE(UUC), trnF(GAA), trnG(GCC), trnG(UCC), trnH(GUG), trnI(CAU)*, trnI(GAU)*, trnK(UUU), trnL(UAA), trnL(UAG), trnL(CAA)*, trnM(CAU), trnM(CAU), trnN(GUU)*, trnP(UGG), trnQ(UUG), trnR(ACG)*, trnR(UCU), trnS(GCU), trnS(GGA), trnS(UGA), trnT(GGU), trnT(UGU), trnV(UAC), trnV(GAC)*, trnW(CCA), trnY(GUA)

\*Gene fully located within the inverted repeats

\*\*Gene partially located within the inverted repeats

**Table 2. Genes containing introns in the chloroplast genomes of *Stenogyne haliakalae* and 11 additional mint species.**

Gene	Location	# Introns	Exon I	Intron I	Exon II	Intron II	Exon III
atpF	LSC	1	143	656	410		
clpP	LSC	2	70	658	291	616	227
ndhA	SSC	1	552	1020	538		
ndhB	IR	1	755	680	776		
petB	LSC	1	5	718	650		
petD	LSC	1	7	728	474		
rpl16	LSC	1	8	907	392		
rpl2	IR	1	390	658	433		
rpoC1	LSC	1	434	736	1,634		
rps12	LSC/IR*	2	113	*	231	537	25
rps16	LSC	1	39	875	226		
trnA-UGC	IR	1	37	807	34		
trnG-UCC	LSC	1	22	690	47		
trnI-GAU	IR	1	34	938	36		
trnK-UUU	LSC	1	36	2,509	34		
trnL-UAA	LSC	1	36	488	49		
trnV-UAC	LSC	1	37	579	36		
ycf3	LSC	2	123	713	229	725	152

\*Trans-spliced

**Table 3. Lengths (bp) of the long single copy region (LSC), short single copy region (SSC), and inverted repeats regions (IR) for the chloroplast genomes of *Stenogyne haliakalae* and 11 additional mint species.**

Species	LSC	IR	SSC	Total
<i>Haplostachys haplostachya</i>	81,755	25,441	17,495	150,132
<i>Haplostachys linearifolia</i>	81,752	25,441	17,495	150,129
<i>Phyllostegia velutina</i>	81,765	25,440	17,496	150,141
<i>Phyllostegia waimeae</i>	81,744	25,448	17,497	150,137
<i>Stachys byzantina</i>	81,245	25,480	17,517	149,722
<i>Stachys chamissonis</i>	81,774	25,464	17,552	150,254
<i>Stachys coccinea</i>	82,171	25,470	17,563	150,674
<i>Stachys sylvatica</i>	81,807	25,414	17,560	150,195
<i>Stenogyne bifida</i>	81,752	25,441	17,495	150,129
<i>Stenogyne haliakalae</i>	81,364	25,436	17,499	149,736
<i>Stenogyne kanehoana</i>	81,739	25,441	17,495	150,116
<i>Stenogyne sessilis</i>	81,743	25,441	17,495	150,120

**Table 4. Sliding window analysis of variability of complete chloroplast genome sequences of three Hawaiian and three *Stachys* taxa (*Stenogyne haliakalae*, *Stenogyne bifida*, *Haplostachys haplostachya*, *Stachys chamissonis*, *Stachys coccinea*, and *Stachys sylvatica*). PICs – Potentially informative characters**

<b>Begin</b>	<b>End</b>	<b># PICs <i>Stachys</i></b>	<b># PICs Hawaiian</b>
1	999	8	1
1000	1999	7	2
2000	2999	6	4
3000	3999	11	1
4000	4999	7	0
5000	5999	5	1
6000	6999	10	1
7000	7999	8	0
8000	8999	6	2
9000	9999	9	0
10000	10999	4	1
11000	11999	3	0
12000	12999	4	0
13000	13999	9	4
14000	14999	4	2
15000	15999	1	0
16000	16999	3	0
17000	17999	6	0
18000	18999	5	3
19000	19999	2	0
20000	20999	3	0
21000	21999	0	0
22000	22999	5	1
23000	23999	1	0
24000	24999	0	0
25000	25999	1	0
26000	26999	4	1
27000	27999	5	3
28000	28999	9	2
29000	29999	4	1
30000	30999	11	1
31000	31999	5	1
32000	32999	3	0
33000	33999	3	1
34000	34999	8	1
35000	35999	1	0

36000	36999	1	0
37000	37999	5	0
38000	38999	0	0
39000	39999	1	0
40000	40999	5	0
41000	41999	3	1
42000	42999	1	0
43000	43999	5	2
44000	44999	6	1
45000	45999	10	0
46000	46999	3	1
47000	47999	7	0
48000	48999	0	0
49000	49999	6	1
50000	50999	4	0
51000	51999	2	0
52000	52999	3	1
53000	53999	5	3
54000	54999	8	3
55000	55999	8	2
56000	56999	2	0
57000	57999	6	1
58000	58999	9	1
59000	59999	4	2
60000	60999	3	3
61000	61999	7	1
62000	62999	2	0
63000	63999	6	1
64000	64999	5	1
65000	65999	6	1
66000	66999	8	0
67000	67999	12	0
68000	68999	6	0
69000	69999	0	0
70000	70999	7	0
71000	71999	5	3
72000	72999	3	0
73000	73999	3	1
74000	74999	4	2
75000	75999	2	1
76000	76999	5	0
77000	77999	5	0
78000	78999	7	0

79000	79999	7	1
80000	80999	4	1
81000	81999	10	0
82000	82999	2	2
83000	83999	0	1
84000	84999	1	0
85000	85999	1	0
86000	86999	2	0
87000	87999	0	0
88000	88999	1	1
89000	89999	0	0
90000	90999	1	0
91000	91999	0	0
92000	92999	1	0
93000	93999	0	0
94000	94999	2	1
95000	95999	1	0
96000	96999	2	0
97000	97999	0	0
98000	98999	0	0
99000	99999	1	0
100000	100999	1	0
101000	101999	1	0
102000	102999	1	0
103000	103999	0	0
104000	104999	5	2
105000	105999	1	0
106000	106999	4	0
107000	107999	9	0
108000	108999	4	0
109000	109999	16	3
110000	110999	6	2
111000	111999	10	1
112000	112999	7	0
113000	113999	8	3
114000	114999	4	0
115000	115999	5	0
116000	116999	6	4
117000	117999	9	4
118000	118999	3	0
119000	119999	5	0
120000	120999	11	2
121000	121999	10	1

122000	122999	7	5
123000	123999	11	2
124000	124999	4	1
<b>Min</b>		0	0
<b>Max</b>		16	5
<b>Total</b>		565	104
<b>Mean</b>		4.52	0.83
<b>Median</b>		4.00	0.00



**Table 5. Comparison of variability of coding genes by exon in complete chloroplast genome sequences of three Hawaiian and three *Stachys* taxa** (*Stenogyne haliakalae*, *Stenogyne bifida*, *Haplostachys haplostachya*, *Stachys chamissonis*, *Stachys coccinea*, and *Stachys sylvatica*).

Note: Exon numbers are defined by position in overall chloroplast genome sequence (not direction of gene) and exons that are completely conserved among these taxa and/or shorter than 100 bp have been excluded. PICs – Potentially informative characters.

Region	Length	#PICs per locus	#PICs per locus	% PICs per locus	% PICs per locus
		<i>Stachys</i>	Hawaiian	<i>Stachys</i>	Hawaiian
psbA	1056	0	2	0.00	0.19
matK	1530	14	5	0.92	0.33
psbK	186	1	0	0.54	0.00
atpA	1524	5	1	0.33	0.07
atpF Exon1	411	1	0	0.24	0.00
atpI	744	1	0	0.13	0.00
rps2	711	1	0	0.14	0.00
rpoC2	4083	16	3	0.39	0.07
rpoC1 Exon1	1635	1	0	0.06	0.00
rpoC1 Exon2	435	1	0	0.23	0.00
rpoB	3213	4	0	0.12	0.00
psbD	1062	4	0	0.38	0.00
psbC	1422	3	1	0.21	0.07
psaB	2205	6	0	0.27	0.00
psaA	2253	2	0	0.09	0.00
rps4	606	1	0	0.17	0.00
ndhJ	477	3	0	0.63	0.00
ndhK	678	3	0	0.44	0.00
atpE	402	2	0	0.50	0.00
atpB	1497	4	0	0.27	0.00
rbcL	1446	12	6	0.83	0.41
accD	1467	3	0	0.20	0.00
ycf4	555	2	0	0.36	0.00
cemA	690	2	1	0.29	0.14
petA	963	2	1	0.21	0.10
rpl33	201	1	0	0.50	0.00
rpl20	387	2	0	0.52	0.00
psbB	1527	9	0	0.59	0.00
petB Exon2	651	2	1	0.31	0.15
rpoA	1014	3	1	0.30	0.10
rpl36	114	2	0	1.75	0.00
rps8	414	1	0	0.24	0.00
rpl14	369	2	0	0.54	0.00

rpl16 Exon1	393	3	0	0.76	0.00
rps3	663	3	0	0.45	0.00
rpl22	465	1	1	0.22	0.22
rps19	279	6	0	2.15	0.00
rpl2 Exon1	434	1	0	0.23	0.00
ycf2	6849	5	2	0.07	0.03
ndhB Exon1	756	1	0	0.13	0.00
rps7	468	2	1	0.43	0.21
rrn23	2811	1	0	0.04	0.00
ndhF	2229	16	0	0.72	0.00
rpl32	177	2	0	1.13	0.00
ccsA	970	3	2	0.31	0.21
ndhD	1521	11	1	0.72	0.07
psaC	244	1	0	0.41	0.00
ndhE	306	1	2	0.33	0.65
ndhG	531	1	0	0.19	0.00
ndhI	507	2	0	0.39	0.00
ndhA Exon1	539	3	0	0.56	0.00
ndhA Exon2	553	3	1	0.54	0.18
ycf1 SSC	4487	43	11	0.96	0.25
<b>Min</b>	114	0	0	0.00	0.00
<b>Max</b>	6849	43	11	2.15	0.65
<b>Total</b>	61110	225	43	23.43	3.45
<b>Mean</b>	1153	4.25	0.81	0.44	0.07
<b>Median</b>	678	2.00	0.00	0.33	0.00

**Table 6. Comparison of variability of introns in complete chloroplast genome sequences of three Hawaiian and three *Stachys* taxa** (*Stenogyne haliakalae*, *Stenogyne bifida*, *Haplostachys haplostachya*, *Stachys chamissonis*, *Stachys coccinea*, and *Stachys sylvatica*). Note: Exon and intron numbers are defined by position in the overall chloroplast genome sequence (not direction of gene). The trnK-UUU intron contains the gene matK. Here trnK-UUU-1 refers to the region between exon 1 and matK, and trnK-UUU-2 refers to the region between matK and exon2. PICs – Potentially informative characters

Region	Length	#PICs per locus	#PICs per locus	% PICs per locus	% PICs per locus
		<i>Stachys</i>	Hawaiian	<i>Stachys</i>	Hawaiian
trnK-UUU - 1	264	5	0	1.89	0.00
trnK-UUU - 2	714	5	0	0.70	0.00
rps16	874	5	1	0.57	0.11
trnG-UCC	689	7	0	1.02	0.00
atpF	655	3	0	0.46	0.00
rpoC1	762	4	1	0.52	0.13
ycf3 - 1	724	0	1	0.00	0.14
ycf3 - 2	712	2	0	0.28	0.00
trnL-UAA	487	4	0	0.82	0.00
trnV-UAC	478	1	1	0.21	0.21
clpP - 1	615	6	0	0.98	0.00
clpP - 2	657	4	0	0.61	0.00
petB	717	2	0	0.28	0.00
petD	727	4	2	0.55	0.28
rpl16	906	6	0	0.66	0.00
rpl2	657	2	2	0.30	0.30
trnI-GAU	937	1	0	0.11	0.00
ndhA	1019	10	7	0.98	0.69
<b>Min</b>	264	0	0	0.00	0.00
<b>Max</b>	1019	10	7	1.89	0.69
<b>Sum</b>	12594	71	15	10.95	1.86
<b>Mean</b>	699.7	3.94	0.83	0.61	0.10
<b>Median</b>	713	4.00	0.00	0.56	0.00

**Table 7. Comparison of variability of intergenic spacers and pseudogenes in complete chloroplast genome sequences of three Hawaiian and three *Stachys* taxa** (*Stenogyne haliakalae*, *Stenogyne bifida*, *Haplostachys haplostachya*, *Stachys chamissonis*, *Stachys coccinea*, and *Stachys sylvatica*). Note: Regions that are completely conserved among these taxa and/or shorter than 100 bp have been excluded. PICs – potentially informative characters

Region	Length	#PICs per locus <i>Stachys</i>	#PICs per locus Hawaiian	% PICs per locus <i>Stachys</i>	% PICs per locus Hawaiian
trnH-GUG-psbA	323	8	1	2.48	0.31
psbA-trnK-UUU	244	2	0	0.82	0.00
trnK-UUU-rps16	643	5	0	0.78	0.00
rps16-trnQ-UUG	828	10	1	1.21	0.12
psbK-psbI	374	7	0	1.87	0.00
psbI-trnS-GCU	121	1	1	0.83	0.83
trnS-GCU-trnG-UCC	694	5	1	0.72	0.14
trnG-UCC-trnR-UCU	160	2	0	1.25	0.00
atpF-atpH	365	5	1	1.37	0.27
atpH-atpI	975	8	5	0.82	0.51
rps2-rpoC2	337	2	0	0.59	0.00
rpoC2-rpoC1	154	1	0	0.65	0.00
rpoB-trnC-GCA	1098	6	4	0.55	0.36
trnC-GCA-petN	436	4	0	0.92	0.00
petN-psbM	509	5	2	0.98	0.39
psbM-trnD-GUC	518	3	0	0.58	0.00
trnE-UUC-trnT-GGU	570	7	1	1.23	0.18
trnT-GGU-psbD	1075	8	1	0.74	0.09
psbC-trnS-UGA	232	3	1	1.29	0.43
trnS-UGA-psbZ	328	3	0	0.91	0.00
psbZ-trnG-GCC	224	2	0	0.89	0.00
trnG-GCC-trnM-CAU	149	1	0	0.67	0.00
psaA-ycf3	730	7	0	0.96	0.00
ycf3-trnS-GGA	316	2	2	0.63	0.63
trnS-GGA-rps4	162	1	0	0.62	0.00
rps4-trnT-UGU	385	3	1	0.78	0.26
trnT-UGU-trnL-UAA	690	9	0	1.30	0.00
trnL-UAA-trnF-GAA	295	3	0	1.02	0.00
trnF-GAA-ndhJ	658	0	1	0.00	0.15
ndhC-trnV-UAC	944	5	0	0.53	0.00
trnV-UAC-trnM-CAU	188	1	0	0.53	0.00
trnM-CAU-atpE	213	2	0	0.94	0.00
atpB-rbcL	801	1	1	0.12	0.12
rbcL-accD	707	7	2	0.99	0.28
accD-psaI	397	1	1	0.25	0.25
psaI-ycf4	434	7	0	1.61	0.00

ycf4-cemA	306	5	1	1.63	0.33
cemA-petA	217	1	1	0.46	0.46
petA-psbJ	1059	9	3	0.85	0.28
psbE-petL	915	6	1	0.66	0.11
petL-petG	174	2	0	1.15	0.00
petG-trnW-CCA	125	1	0	0.80	0.00
trnP-UGG-psaJ	270	2	1	0.74	0.37
psaJ-rpl33	478	5	0	1.05	0.00
rpl33-rps18	146	0	1	0.00	0.68
rps18-rpl20	216	5	0	2.31	0.00
rpl20-rps12	739	10	0	1.35	0.00
psbB-psbT	184	1	1	0.54	0.54
psbH-petB	124	1	0	0.81	0.00
petB-petD	194	1	0	0.52	0.00
infA-rps8	123	1	0	0.81	0.00
rps8-rpl14	173	3	0	1.73	0.00
rpl14-rpl16	126	1	0	0.79	0.00
rpl16-rps3	141	1	1	0.71	0.71
rps12-3'end-trnV-GAC	1448	3	0	0.21	0.00
trnA-UGC-rrn23	199	1	0	0.50	0.00
rrn4.5-rrn5	224	2	1	0.89	0.45
rrn5-trnR-ACG	234	2	1	0.85	0.43
trnR-ACG-trnN-GUU	569	2	0	0.35	0.00
ycf1 Truncated	1059	1	0	0.09	0.00
ndhF-rpl32	435	5	2	1.15	0.46
rpl32-trnL-UAG	737	13	2	1.76	0.27
ccsA-ndhD	180	4	0	2.22	0.00
ndhD-psaC	105	1	0	0.95	0.00
psaC-ndhE	251	5	0	1.99	0.00
ndhG-ndhI	375	6	0	1.60	0.00
rps15-ycf1 SSC	389	3	0	0.77	0.00
<b>Min</b>	105	0	0	0.00	0.00
<b>Max</b>	1448	13	5	2.48	0.83
<b>Sum</b>	29192	250	43	62.72	10.44
<b>Mean</b>	435.7	3.73	0.64	0.94	0.16
<b>Median</b>	328.0	3.00	0.00	0.82	0.00

**Table 8. Tailed primers used for multiplex amplification and targeted re-sequencing in mints.** Sequences complimentary to the Illumina sequencing adapters were appended to the end of each primer (Forward: 5' TCGTCGGCAGCGTCAGATGTGTATAAGAGACAG-[locus specific sequence] and Reverse: 5' GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAG-[locus specific sequence]).

Primer	Sequence 5' – 3'	Tm	Loci in region (excluding priming sites)
Mint204F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CGC CCC TCT ACT ATA ATG AAT GA	69.3	trnH-GUG_psbA
Mint204R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GAC AGG ATC CAG AAA AAG AAA GA	68.1	
Mint771F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CCA TGA GCG GCT ACG ATA TT	70.3	psbA
Mint771R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCA GGC TGA GCA CAA CAT TCT	70.2	
Mint867F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG GGA ACC ATG CAT AGC ACT GA	70.4	psbA
Mint867R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GTA CCC AAT CGG TCA AGG AAG	69.1	
Mint1170F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CAC TCA CGA CCC ATG TAA CAA	70.1	psbA
Mint1170R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GTG AGC CTG TTT CTG GAT CTC T	69.5	
Mint1939F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG TCA AAA CAA AAG TTG AAT ACT CAG TTG	67.6	trnK-UUU Intron1, matK
Mint1939R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCG GAT TTG GTA TTT GGA TAT GA	68.3	
Mint3998F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG GCT TCC CGT ATC AGG CAC T	71.2	trnK-UUU Intron2
Mint3998R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GTC GAA TTC TTG GAA CGG AAC	68.6	
Mint4546F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CAG AAT TGT CAA AAT GTA TAG AGC A	68.2	trnK-UUU Exon2_rps16 Exon1
Mint4546R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCA TCG TGA TAA GCG ATC TGG	69.2	
Mint4714F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG GAC CCA GAT CGC TTA TCA CG	70.1	trnK-UUU Exon2_rps16 Exon1
Mint4714R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GTG TCA ATC CAA GAC AAT TTT GAA	67.8	
Mint5533F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG TCC GAT CCA GTT ATT GAG ACG	69.0	rps16 Intron
Mint5533R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCG GGA ATC GAC TGT CCA TAG	69.7	
Mint5985F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG GCC CCC GAG AAA TGA ATT A	69.9	rps16 Intron
Mint5985R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GTA GAA AGC AAC GTG CGA CTT	69.3	

Mint6787F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG TTC CAA TTT TGC ATT CGA GTC	68.5	rps16 Exon2_trnQ-UUG
Mint6787R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GAA AAG GGT GAG TGG GTA GGA	69.7	
Mint7334F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG TCA AAA ACG CAA CCA AAA TG	68.4	trnQ-UUG_psbK, psbK
Mint7334R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GTG GCA TAA CAT CCA CGA TTG	68.8	
Mint8052F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG GGT TTC CTT GGG TTT GGT TA	69.9	psbI-trnS-GCU, trnS-GCU
Mint8052R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GGG AGA GAT GGC TGA GTG GAC	70.3	
Mint8691F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CCG AAC TCA AAA ATA AAC TGT CG	68.7	trnS-GCU_trnG-UCC Exon1
Mint8691R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCA AAA CGA GAA CGT TGC ACT	69.7	
Mint8791F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG ATG AAG CCT CTT TCC CGA A	69.8	trnS-GCU_trnG-UCC Exon1,
Mint8791R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCG GTT ACT AGA ACG AAT CAC ACT TT	68.9	trnG-UCC Exon1
Mint10381F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG TGC GCC AAT TCC AAT TTT A	69.0	atpA
Mint10381R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GTC AAT CGG GAG ATG TTT CG	68.7	
Mint10531F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CAT TAA TGG CGG GTC TGA TT	69.9	atpA
Mint10531R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCT TTT GGA AAG AGC CGC TAA	69.6	
Mint11293F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CTG AGC AAT TCT TCC TGT TGC	69.9	atpA
Mint11293R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GGG CGA TGG TAT TGC TCG TAT	69.8	
Mint13441F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CAG CAG CAA TAA CGG AAG C	70.3	atpH, atpH_atpI
Mint13441R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCG AAG TCG TTC TGA TGA TTC AA	68.8	
Mint13731F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG AGC CAC GAC GAT ATG AAA GG	69.4	atpH_atpI
Mint13731R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GAA AGT GGA TTG GTT GTC GAA	68.7	
Mint14054F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG GAT TGA TCT AAG TTC ATG CAA TTT TT	67.8	atpH_atpI
Mint14054R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GTT GTC CAC TTA ATA TCC TAC CTT TCC	67.9	
Mint16824F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG TGA AAT CAA GAC CTT TGA TGT TAT T	67.7	rpoC2
Mint16824R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GGA GGG TTG GAA CGA ACG TAT	69.8	
Mint17255F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CCG GAG TGG CCA AAT AAG T	70.8	rpoC2
Mint17255R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GTT GGT ATT TTC TCC ATC CCA AT	68.3	



Mint17928F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG TGG ATT TCA GCA AGT CGA TTC	68.9	rpoC2
Mint17928R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCC TTT ATG GAA ATG GGA AAC C	68.8	
Mint20423F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG ATG CCA TTC CGA AGT GAT CT	69.6	rpoC2, rpoC2_rpoC1 Exon1
Mint20423R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCG CGA ATC AAG ATC GAG AAC	69.3	
Mint20572F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CGA TTT GAC AAT GGG TTT GA	69.4	rpoC2_rpoC1 Exon1,
Mint20572R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GAA GCC ATA CAG GGG TTT TCC	69.4	rpoC1 Exon1
Mint21910F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG GAG CTC ATT AAT TTA CCC CCA TC	69.0	rpoC1 Exon1
Mint21910R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GGG GAA TGA ATG GGA AGA TCA	69.2	
Mint22486F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG TCA AGT TAC CAG TGA AGA CTA AGC A	69.0	rpoC1 Intron
Mint22486R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GGA TTC ATC ATT CGA AGG GAA GT	68.8	
Mint23294F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG AAA TTC GAC TCC GCA TTG TT	69.2	rpoC1 Exon2
Mint23294R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GGC CCA ATG GAG AGA TAG TCG	69.7	
Mint23536F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG AAT CCA ATT CGG AGC TGT TG	69.1	rpoB
Mint23536R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GTG GTC AAG TCA TAA AGT CAA ATA AA	67.2	
Mint24088F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG GGC GCT ATT CGA TAA TGT CTG	69.3	rpoB
Mint24088R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GGG AAG ACA CGG AWA CAA AGG	69.3	
Mint24204F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CAA CAG GTC TTC CAT CTT GC	69.9	rpoB
Mint24204R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCC GGG TTA TTG ATG TGA GGT	70.0	
Mint27582F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CTG ATT GGT TCA GTC TCA GTT TTT	69.1	rpoB_trnC-GCA
Mint27582R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GGC GAG AGA ATG TTT TTA GCA TTG	68.4	
Mint27884F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG GCA AAT CCT TTT TCC CCA GT	70.1	trnC-GCA, trnC-GCA_petN
Mint27884R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GAA CGG GGC TTC ACA ATC TTT	69.5	
Mint28672F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG AAG AGA TTC GGA TGA TTG GAA A	68.4	petN_psbM
Mint28672R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GCG AGC GCA CTA TAA TCA GCA	69.9	
Mint28881F	TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG GCT GAT TAT AGT GCG CTC GTT	70.0	petN_psbM, psbM
Mint28881R	GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GTC CTA CCG CCT TTC TGC TTA	69.7	

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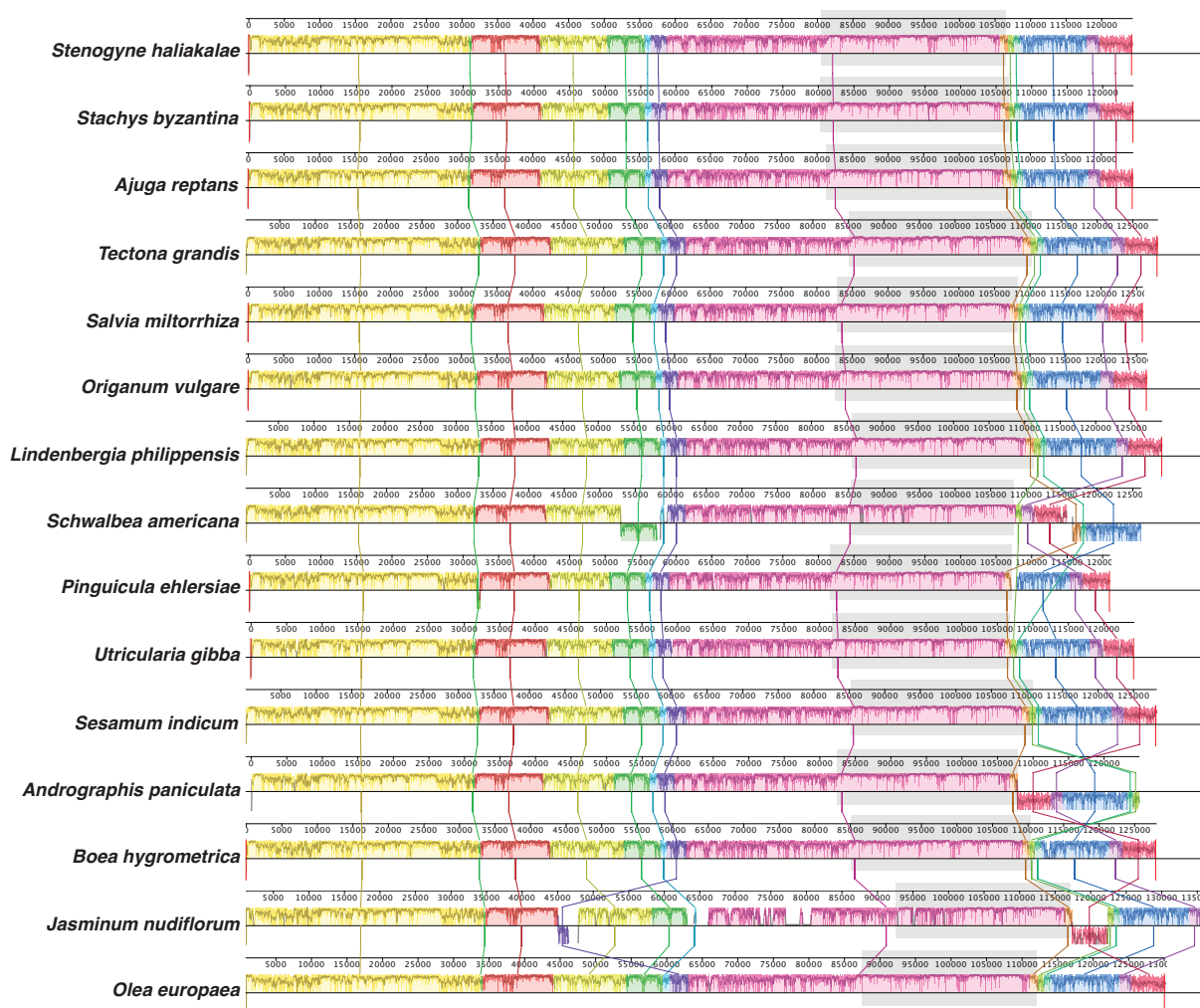
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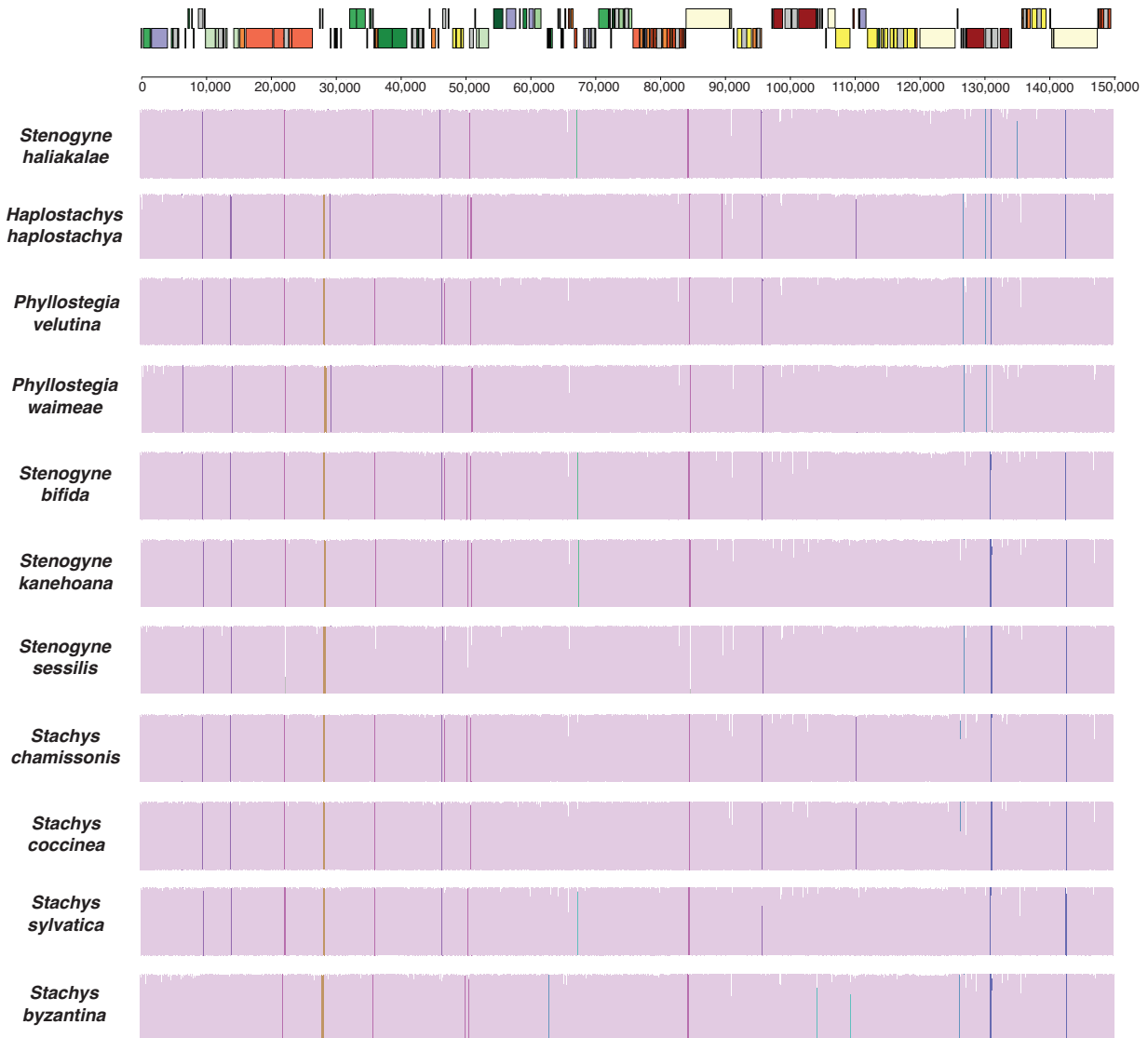


## FIGURES

**Figure 1. Comparison of structure and similarity among 15 complete chloroplast genomes from the order Lamiales.** *Cistanche deserticola* (102,657 bp) and *Epifagus virginiana* (70,028 bp), both members of the Orobanchaceae, are not considered here because they are parasitic and lack chlorophyll, thus demonstrating largely reduced chloroplast genomes. Blocks with the same color represent homologous regions free of internal structural changes for that subset of taxa, and those above the centerline for each taxon are in the same orientation as in *Stenogyne haliakalae*, whereas those below the line are in the reverse direction. Within each block a similarity profile for the region is plotted. Areas outside of blocks are presumed to represent lineage-specific regions of the chloroplast genome. One copy of the inverted repeat has been trimmed so that homology of the remaining repeat (area shaded in light gray) can be shown.

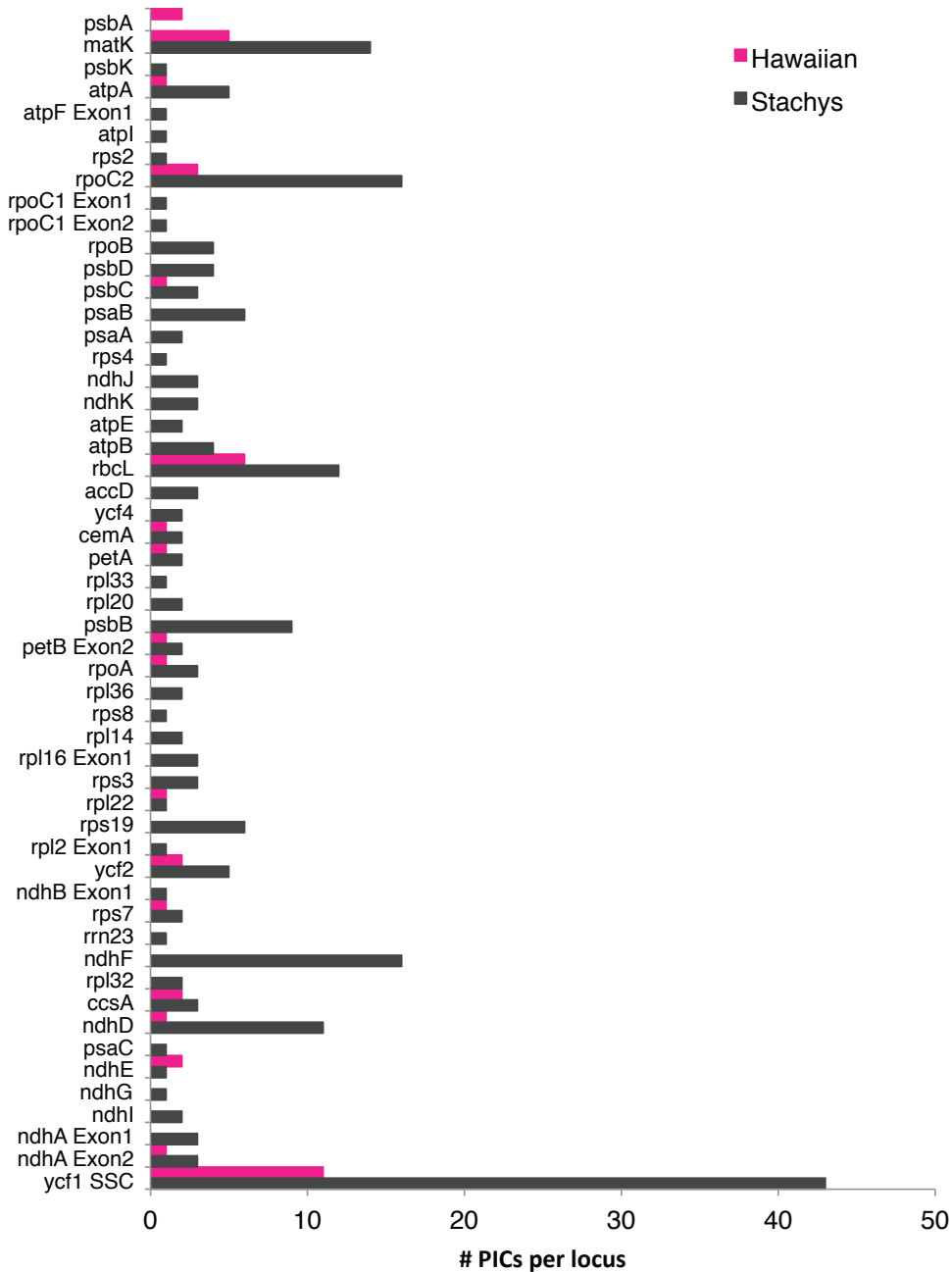


**Figure 2. Conservation among 11 complete mint chloroplast genomes.** The sequence for *Haplostachys linearifolia* was excluded due to missing data. A physical map is given at the top to show gene content and organization (see Figure 2 in [1] for gene names and products). In the lower panels, regions of the genome are represented by bars, and those that are conserved among all 11 species are colored mauve, whereas those that are conserved among subsets of the taxa have different colors. The height of the bar shows the degree of similarity.

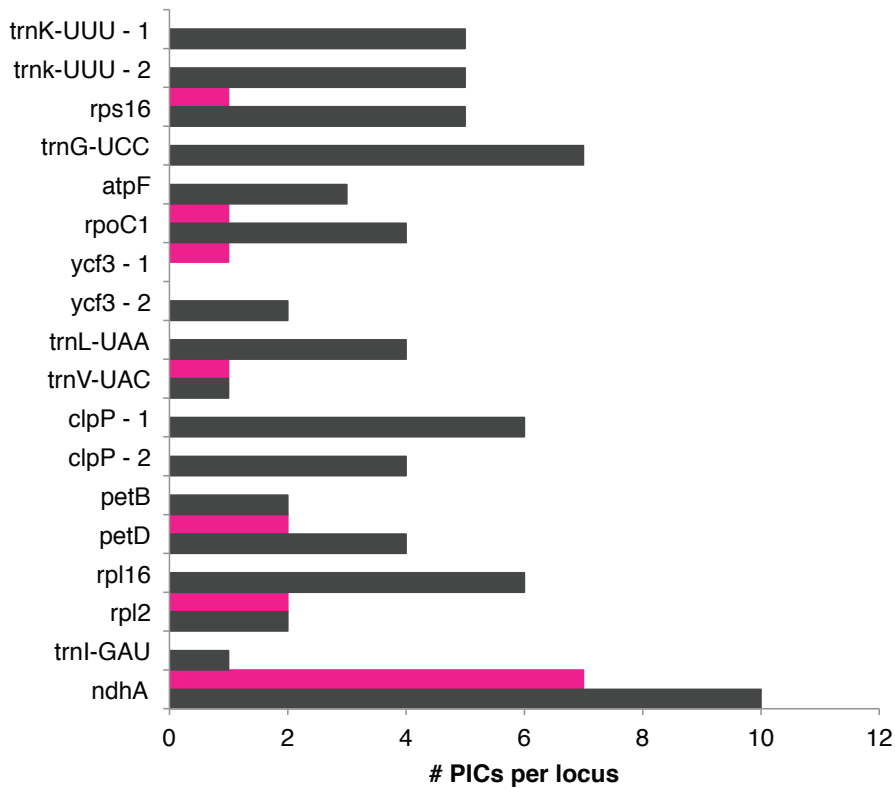


**Figure 3. Comparison of the number of potentially informative characters (PICs) per locus in a) protein coding regions, b) introns, and c) intergenic spacers and pseudogenes for chloroplast genomes of three Hawaiian and three *Stachys* taxa: *Stenogyne haliakalae*, *Stenogyne bifida*, *Haplostachys Haplostachya*, *Stachys chamissonis*, *Stachys coccinea*, and *Stachys sylvatica*. Regions that were completely conserved among these species and/or that are shorter than 100 bp are not shown. Pink = three Hawaiian taxa; Black = three *Stachys* taxa.**

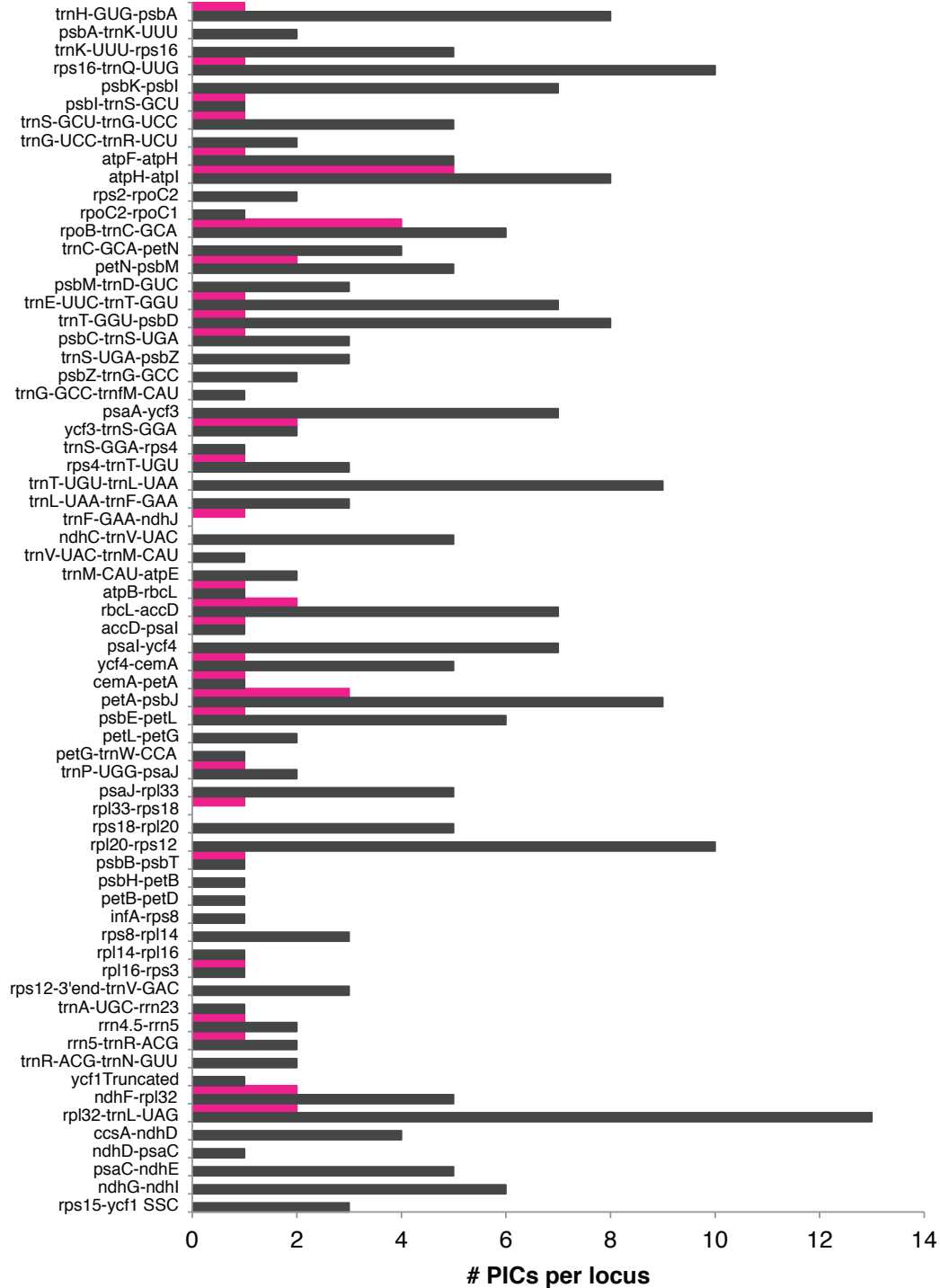
**3a)**



3b)

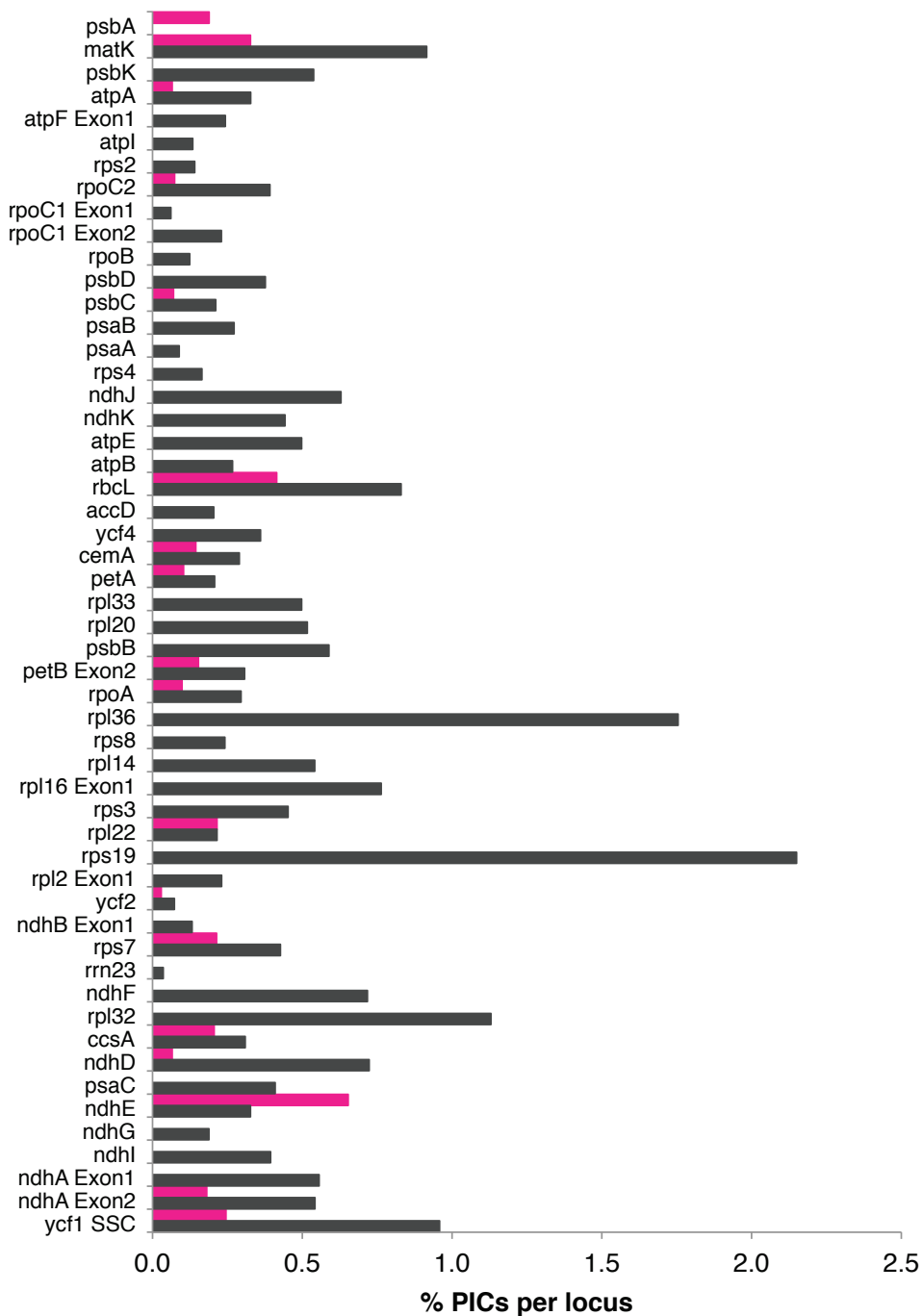


3c)

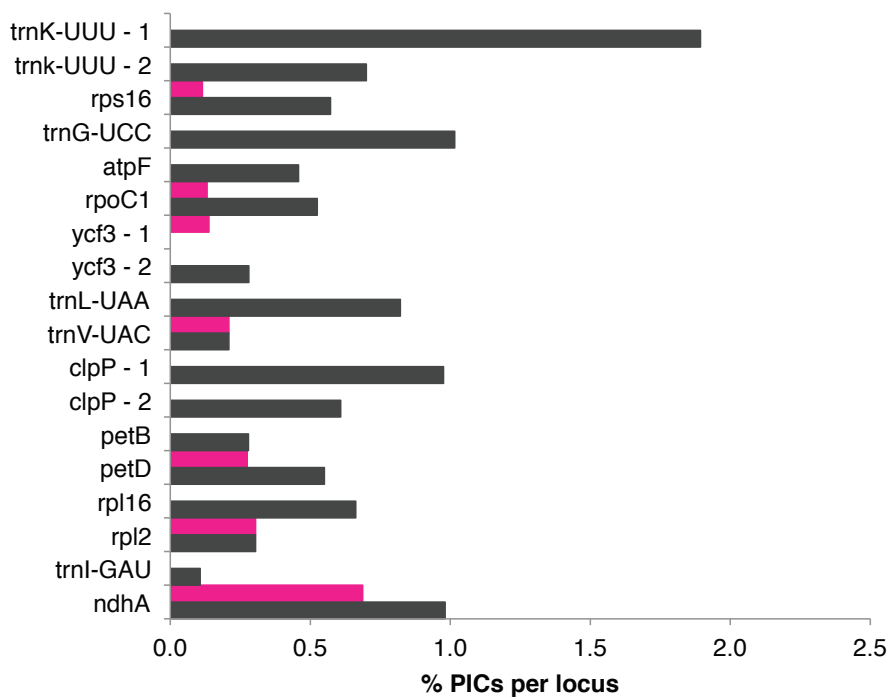


**Figure 4. Comparison of the % PICs (potentially informative characters) per locus in a) protein coding regions, b) introns, and c) intergenic spacers and pseudogenes for chloroplast genomes of three Hawaiian and three *Stachys* taxa: *Stenogyne haliakalae*, *Stenogyne bifida*, *Haplostachys haplostachya*, *Stachys chamissonis*, *Stachys coccinea*, and *Stachys sylvatica*. Regions that were completely conserved among these species or that are shorter than 100 bp are not shown. Pink = three Hawaiian taxa; Black = three *Stachys* taxa.**

**4a)**



4b)





4c)

